

Formation of Acyclic α - and γ -Ketols and 12-Oxophytodienoic Acid from Linolenic Acid 13-Hydroperoxide by a Flax Enzyme Preparation. Evidence for a Single Enzyme Leading to a Common Allene Epoxide Intermediate

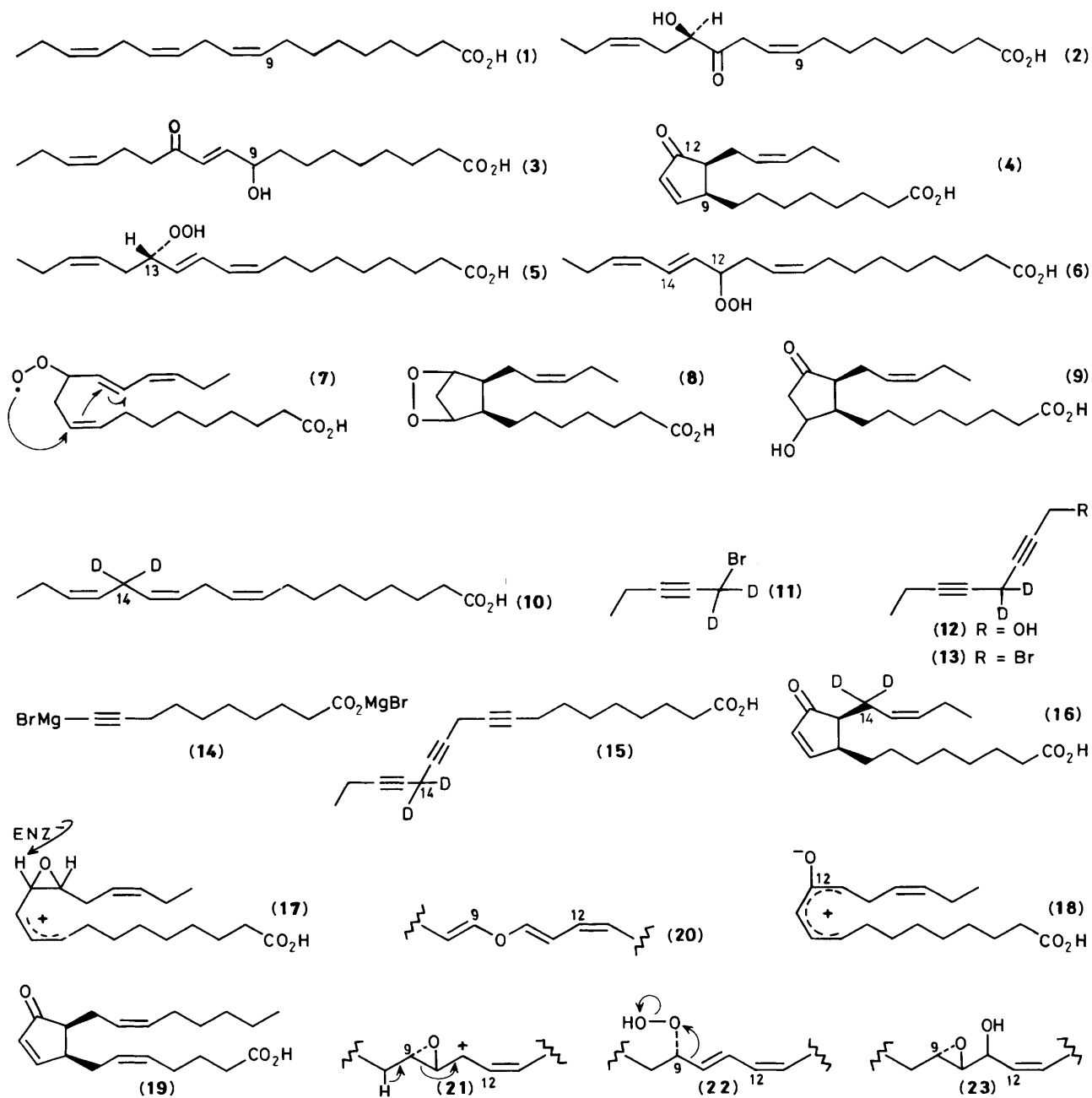
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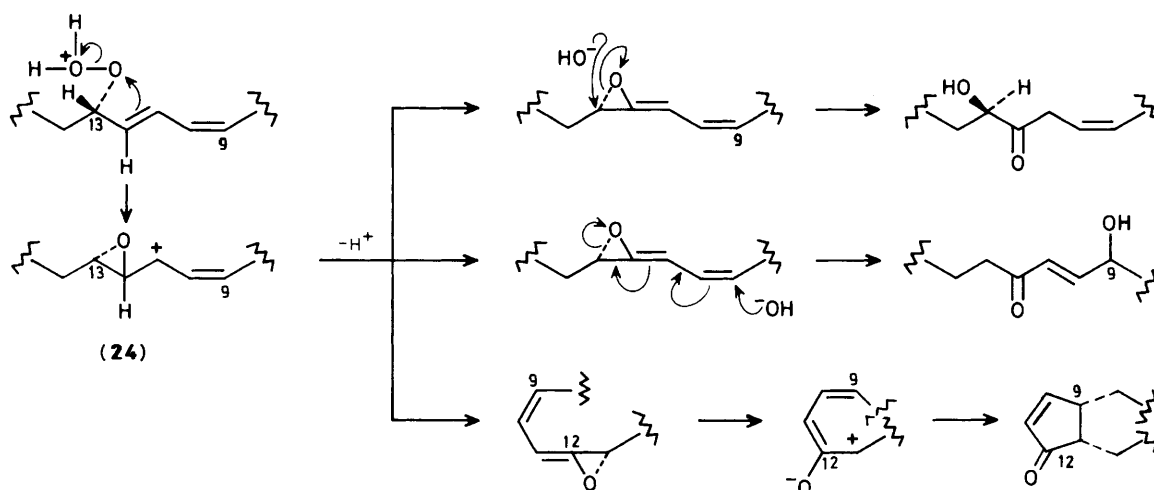
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Experiments with $[14\text{-}^2\text{H}_2]$ linolenic acid confirm the requirement for a 13-hydroperoxide for 12-oxophytodienoic acid (12-oxoPDA) formation by a flax enzyme preparation; an allene epoxide intermediate is capable of explaining the formation of the acyclic α - and γ -ketols as well as 12-oxoPDA, suggesting the involvement of a single enzyme.

Zimmerman *et al.* have described an enzyme preparation from flax seed capable of converting linolenic acid (1) into the α -ketol (2), the γ -ketol (3), and the prostaglandin (PGA₁)-like 12-oxophytodienoic acid (12-oxoPDA) (4).¹ This system is of

considerable importance to plant physiology, and to food chemistry, as it is widespread among higher plants.² The precursor to (4) is considered to be the 13-hydroperoxide (5) derived from linolenic acid (1).¹ Were the biosynthesis of





Scheme 1. Allene-epoxide mechanism for the formation of the α - and γ -ketols, and 12-oxoPDA from linolenic acid 13-hydroperoxide and flax enzyme.

12-oxoPDA to follow a pathway resembling that of mammalian prostaglandins, a 12-hydroperoxide (6) would be involved in the pathway leading to (4) via (7), (8), and (9).³ Since there is some evidence that a 12-hydroperoxide may be formed by flax seed lipoxygenase,⁴ we administered h.p.l.c. pure [1-¹⁴C]-13-hydroperoxyoctadeca-(9Z),(11E),(15Z)-trienoic acid (5) to the flax seed cyclase enzyme. Although satisfactory incorporation into 12-oxoPDA was eventually obtained, there were initial difficulties which led to the experiment below. This demonstrates in a different way that the 13- and not the 12-hydroperoxide is involved.

[14-²H₂]Linolenic acid (10) was synthesised as follows. But-1-yne (as its Grignard complex) was converted into [1-²H₂]pentynol by treatment with deuterioformaldehyde (made by deuteration of dibromomethane, conversion into [2H₂]methylene diacetate, and hydrolysis).⁵ [1-²H₂]Pentynol was converted (PBr₃) into the bromide (11) and coupled (copper catalyst)⁶ with the Grignard reagent from tetrahydropyranyl-protected prop-2-ynyl alcohol to give (12), itself converted into the bromide (13). The latter was coupled under copper cyanide catalysis with the acetylenic bis-Grignard (14). Triyne-acid (15) was esterified (CH₂N₂), reduced with diisopentylborane,⁷ and hydrolysed to give [14-²H₂]linolenic acid (10). Treatment with flax seed cyclase enzyme,¹ followed by chromatographic work-up, gave 12-oxoPDA as its methyl ester (CH₂N₂). This retained both deuterium atoms (*M*⁺ 306 + 2, *M*⁺ - OMe 275 + 2 etc.) (16). The ¹H n.m.r. spectrum of (16), when compared with that of the undeuteriated material, showed that both deuteriums were still located on the 14-methylene (16). Thus the multiplet at δ 2.51 (1H, 14-H_a) and the ddd at 2.15 (1H, 14-H_b) were not present: the erstwhile ddd at 2.44 (1H, 13-H) now appeared as a clean doublet and the olefinic multiplet at 5.40 (H-15 and H-16) was simplified. It is clear that the 13-, not the 12-, hydroperoxide must be the precursor of 12-oxoPDA, with oxygen migration from C-13 to C-12 occurring in the biosynthesis.

Zimmerman⁸ has proposed that the cyclase enzyme leads to an epoxy-carbonium ion (17) from which enzymic loss of a proton leads to zwitter-ion (18) which undergoes cyclisation to give (4). Corey has lately proposed a similar antarafacial pericyclic closure to explain the formation of preclavulone A (19),⁹ produced by treating arachidonic acid with an homogenate of the coral *Clavularia viridis*, and has pointed out a chemical analogy for the process.^{10,11} This compound is

also a *cis*-compound unlike the mammalian prostaglandin series.

The three products from the flax enzyme system and linolenic acid can now be assembled into a single mechanistic scheme in which the central feature is the formation of a protonated allene epoxide (24) (Scheme 1), deprotonated to the allene epoxide itself. One branch leads to the α -ketol,¹² one to the γ -ketol,¹³ and a third to 12-oxoPDA as shown. It has not been possible to resolve the flax enzyme preparation into an isomerase and a cyclase enzyme,¹⁴ and the present chemical knowledge leads us the view that these functions merge and that a single enzyme is involved in initiating these processes.

The potato enzyme system^{15,16} which operates on the hydroperoxides of linoleic and linolenic acid has some similarities. Formation of the colnelaic/colnelenic system (20) at pH 7–9 has earlier been shown by us to involve (21) though it could also be represented as a less likely Baeyer–Villiger C \rightarrow O type shift (22). However, at pH 5–7 it may be significant that the same enzyme leads to (21) in hydroxy-trapped form (23).¹⁷

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